



ORIGINAL ARTICLE

# High frequency of methicillin-susceptible and methicillin-resistant *Staphylococcus aureus* in children under 1 year old with skin and soft tissue infections<sup>☆</sup>



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## KEYWORDS

*Staphylococcus aureus*;  
MSSA;  
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Pediatric;  
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Molecular biology

## Abstract

**Objective:** *Staphylococcus aureus* is responsible for a large number of infections in pediatric population; however, information about the behavior of such infections in this population is limited. The aim of the study was to describe the clinical, epidemiological, and molecular characteristics of infections caused by methicillin-susceptible and resistant *S. aureus* (MSSA–MRSA) in a pediatric population.

**Method:** A cross-sectional descriptive study in patients from birth to 14 years of age from three high-complexity institutions was conducted (2008–2010). All patients infected with methicillin-resistant *S. aureus* and a representative sample of patients infected with methicillin-susceptible *S. aureus* were included. Clinical and epidemiological information was obtained from medical records and molecular characterization included *spa* typing, pulsed-field gel electrophoresis (PFGE), and multilocus sequence typing (MLST). In addition, staphylococcal cassette chromosome *mec* (SCC*mec*) and virulence factor genes were detected.

**Results:** A total of 182 patients, 65 with methicillin-susceptible *S. aureus* infections and 117 with methicillin-resistant *S. aureus* infections, were included in the study; 41.4% of the patients being under 1 year. The most frequent infections were of the skin and soft tissues. Backgrounds such as having stayed in day care centers and previous use of antibiotics were more common in patients with methicillin-resistant *S. aureus* infections ( $p \leq 0.05$ ). Sixteen clonal complexes were identified and methicillin-susceptible *S. aureus* strains were more diverse. The most common cassette was staphylococcal cassette chromosome *mec* IVc (70.8%), which was linked to Pantón–Valentine leukocidin (*pvl*).

**Conclusions:** In contrast with other locations, a prevalence of infections in children under 1 year of age in the city could be observed; this emphasizes the importance of epidemiological knowledge at the local level.

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**PALAVRAS-CHAVE**

*Staphylococcus aureus*;  
MSSA;  
MRSA;  
Pediatria;  
Epidemiologia;  
Biologia molecular

**Alta frequência de *S. aureus* (MSSA-MRSA) em crianças com menos de um ano de idade com infecções de pele e do tecido mole****Resumo**

**Objetivo:** O *Staphylococcus aureus* é responsável por um grande número de infecções na população pediátrica; contudo, as informações sobre o comportamento dessas infecções nessa população são limitadas. O objetivo do estudo foi descrever as características clínicas, epidemiológicas e moleculares de infecções causadas por *Staphylococcus aureus* suscetíveis e resistentes à metilina (MSSA-MRSA) em uma população pediátrica.

**Método:** Um estudo transversal descritivo foi realizado em pacientes entre 0 e 14 anos de idade de três instituições de alta complexidade (2008-2010). Todos os pacientes infectados com *S. aureus* resistentes à metilina e uma amostra representativa de pacientes infectados com *S. aureus* suscetíveis à metilina foram incluídos. As informações clínicas e epidemiológicas foram obtidas de prontuários médicos, e a caracterização molecular incluiu tipagem *spa*, Eletroforese em Gel de Campo Pulsado (PFGE) e Tipagem de sequências multilocus (MLST). Além disso, o Cassete Cromossômico Estafilocócico *mec* (SCC*mec*) e genes de fatores de virulência foram detectados.

**Resultados:** 182 pacientes, 65 com infecções por *S. aureus* suscetíveis à metilina e 117 com infecções por *S. aureus* resistentes à metilina, foram incluídos no estudo; 41,4% dos pacientes com menos de um ano de idade. As infecções mais frequentes foram da pele e dos tecidos moles. Os históricos como internações em centros de atendimento e o uso prévio de antibióticos foram mais comuns em pacientes com infecções por *S. aureus* resistentes à metilina ( $p \leq 0,05$ ). Dezesesseis complexos clonais foram identificados, e as cepas de *S. aureus* suscetíveis à metilina foram mais diversificadas. O cassete mais comum foi o Cassete Cromossômico Estafilocócico *mec* IVc (70,8%), relacionado à leucocidina de panton-valentine (*pvl*).

**Conclusões:** Em comparação a outros locais, observamos uma prevalência de infecções em crianças com menos de um ano de idade na cidade; o que enfatiza a importância de conhecer a epidemiologia em nível local.

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**Introduction**

The complex situation of worldwide *Staphylococcus aureus* resistance to methicillin has led to its current management, representing a priority for the World Health Organization (WHO) and a challenge for human public health in different regions; however, both methicillin-susceptible and resistant *S. aureus* (MSSA and MRSA) strains possess a virulence and pathogenic capacity that allows them to reach high infection rates.

Some population groups are more susceptible to *S. aureus* infections; in particular, the pediatric population has less effective immunological function, persistently high bacterial colonization rates, poor hygiene habits, and constant exposure to school environments that favor the acquisition of infection and the dissemination of the microorganism.<sup>1,2</sup> The epidemiology of *S. aureus* infections in this population is diverse, and the frequencies of MRSA infection differ between geographic regions, ranging from 6%<sup>3</sup> to 69.9%<sup>4</sup>; in Colombia, frequencies up to 47.4% have been found.<sup>5</sup>

In Medellín, although *S. aureus* is one of the main agents responsible for infections in the pediatric population, both in hospitals and in the community, there is little information on the characteristics of the infections caused by this microorganism in this population. This study aims to describe the clinical, epidemiological, and molecular characteristics of *S. aureus* infections (MSSA-MRSA) in a pediatric population of the city.

**Methods****Study population**

An observational cross-sectional study was conducted from February 2008 to June 2010, at three tertiary care hospitals from Medellín, the second largest city in Colombia. Patients between birth and 14 years infected with *S. aureus* (MSSA-MRSA) were recruited prospectively; only the first isolate from each individual was evaluated. All patients with MRSA isolates obtained during the time of the study were included, and considering that the prevalence of MSSA is higher, a sample was defined. The sample size was calculated based on the MSSA prevalence during 2007 within each institution, which numbered 65 isolates. The MSSA isolates included were randomly selected each month, from February 2008 to June 2010, using a table of random numbers according to records of each participating institution.

The research protocol and informed consent (signed by parents or guardians) were approved by the Bioethics Committee for Human Research of the University Research Center at Universidad de Antioquia (approval no. 0841150) and by the bioethics committee of each hospital.

**Clinical and epidemiological data**

Clinical and epidemiological data for each patient were obtained from medical records. Information included

clinical and demographic characteristics, antimicrobial use, risk factors, co-morbidities, type of infection, treatment, length of hospital stay, and outcome. According to the Centers for Disease Control and Prevention (CDC) criteria, an infection was considered present on admission (POA) if the date of the event occurred on the day of admission to an inpatient location, two days before admission, or the calendar day after admission. An infection was considered health care-associated (HAI) if the date of event occurred on or after the third calendar day of admission to an inpatient location. Patients with surgical site infections and ventilator-associated event were excluded.<sup>6</sup>

### Identification and antibiotic susceptibility

Identification of *S. aureus* was conducted by standard laboratory methods based on colony morphology in sheep blood agar and positive catalase and coagulase tests. Antibiotic susceptibilities of *S. aureus* isolates were assessed in accordance with Clinical Laboratory Standards Institute guidelines (CLSI, 2009) using the VITEK<sup>®</sup> 2 system (BioMérieux, Inc., NC, EUA). The antibiotics tested included clindamycin, erythromycin, gentamycin, linezolid, moxifloxacin, oxacillin, rifampicin, tetracycline, tigecycline, trimethoprim-sulfamethoxazole, and vancomycin. The *S. aureus* strain ATCC 29213 was used for quality control.

### Polymerase chain reaction (PCR) confirmation of *S. aureus* and methicillin resistance

The presence of the species-specific *nuc* and *femA* genes as well as the *mecA* gene were verified by PCR, as previously described.<sup>7,8</sup>

### Molecular typing: *spa* typing (*spa*), multilocus sequence typing (MLST) and pulsed field gel electrophoresis (PFGE)

In all isolates, the polymorphic X region of the protein A gene (*spa*) was amplified and sequenced as previously described.<sup>9</sup> Corresponding *spa* types were assigned using eGenomics software<sup>9,10</sup> and Ridom *spa*-types were subsequently assigned using the *spa* typing website (<http://www.spaserver.ridom.de/>) developed by Ridom GmbH and curated by SeqNet.org (<http://www.SeqNet.org/>).<sup>11</sup> MLST was performed on a subset of ten isolates representing the more frequent *spa* types, using the methodology described by Enright et al.<sup>12</sup> Allele numbers and sequence types (ST) were assigned using the database maintained at <http://saureus.mlst.net/while> clonal complexes (CC) were inferred using eBURST analysis.<sup>13</sup> Clonal complexes for all remaining strains were inferred by *spa* repeat pattern analysis,<sup>10</sup> or by referring to the Ridom *spa* server website.

PFGE, following *Sma*I digestion, was performed according to a protocol described elsewhere.<sup>14</sup> DNA fragment patterns were normalized using *S. aureus* strain NCTC 8325. Cluster analysis was performed using the Dice coefficient in BioNumerics software (BioNumerics<sup>®</sup>, software version 6.0, Belgium). Dendrograms were generated by the unweighted

pair group method using average linkages (UPGMA). Similarity cut-offs of 80% and 95% were used to define types and subtypes, respectively.<sup>14</sup>

### SCCmec typing

For MRSA isolates, SCCmec types and subtypes were determined using sets of multiplex PCR reactions, as previously described.<sup>15,16</sup> MRSA strains were included as positive controls for SCCmec types and subtypes.

### Detection of staphylococcal virulence factors

All isolates were screened for the genes encoding staphylococcal enterotoxins (*sea*, *seb*, *sec*, *sed*, *see*), toxic shock syndrome toxin 1 (*tst*), and exfoliative toxins *a* and *b* (*eta*, *etb*), using the protocols and primers described by Mehrotra et al.<sup>8</sup> The *lukS/F-PV* genes, encoding Pantone-Valentine leukocidin (PVL); and the *arcA* gene, associated arginine catabolic mobile element (ACME), were also assessed by PCR.<sup>17,18</sup>

### Statistical analyses

Comparisons of clinical, epidemiological, and molecular characteristics were conducted between MSSA and MRSA infected patients, and among the different groups of infections obtained after applying CDC criteria.

Categorical variables were compared using the chi-squared test or Fisher's exact test; Student's *t*- and Mann-Whitney *U* tests were used for continuous variables. *p* values  $\leq 0.05$  were considered statistically significant. Statistical analyses were carried out using the software package in SPSS<sup>®</sup> (IBM SPSS Statistics for Windows, version 21.0, NY, USA).

## Results

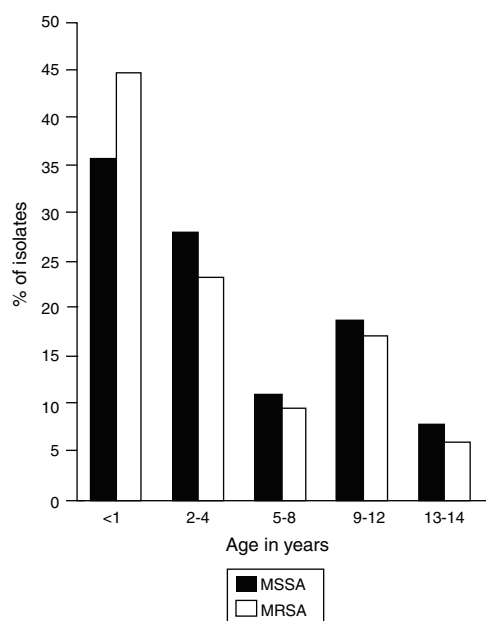
### Clinical and epidemiological data

There were 182 pediatric patients with *S. aureus* infections included; of these, 65 had MSSA infections and 117 had MRSA infections; 119 patients came from hospital A, 51 from hospital B, and 12 from hospital C.

In the selected study population, the ratio of boys and girls was 2:1 and the median age was 2 years; however, 41.2% ( $n = 75$ ) of the infections occurred in patients between birth and 1 year of age, in which a high frequency of MRSA and MSSA infections was observed (Fig. 1).

According to the CDC criteria, 62.7% ( $n = 101$ ) of the infections were POA, and 37.3% ( $n = 60$ ) corresponded to HAI ( $p = 0.391$ ). Infection type showed statistically significant differences by hospital. In hospitals A and B there was a predominance of POA infections; in hospital C there was a predominance of HAI ( $p = 0.029$ ). Twenty-one (11.5%) surgical site infections were identified, which were not included in this classification.

The clinical-epidemiological characteristics of the patients evaluated in the study are described in Table 1. In general, a previous history of hospitalization was the most frequent antecedent among patients, with 51.65% ( $n = 94$ );



**Figure 1** Frequency of infection by age group. MSSA, methicillin-susceptible *Staphylococcus aureus* ( $n=65$ ); MRSA, methicillin-resistant *Staphylococcus aureus* ( $n=117$ ). The figure shows the age distribution of patients infected by MSSA and MRSA strains. A higher frequency of infections is observed in children under 1 year.

furthermore, having stayed in day care centers (MRSA 11.1%,  $n=13$  vs. MSSA 1.5%,  $n=1$ ;  $p=0.020$ ) and previous use of antibiotics (MRSA 55%,  $n=60$  vs. MSSA 32.8%,  $n=20$ ;  $p=0.005$ ) was the most frequent antecedent in patients with MRSA infections.

In HAI, a previous history of intensive care unit (ICU) stay (MRSA 21.4%,  $n=9$  vs. MSSA 0%;  $p=0.047$ ) and prior antibiotic use in the last six months (MRSA 64.1%,  $n=25$  vs. MSSA 29.4%,  $n=5$ ;  $p=0.017$ ) were more frequent in patients with MRSA infections.

However, the hospital stay was longer in patients with HAI, with a significant difference in favor of patients with MRSA infections (MRSA 0–434 days,  $M_e=14$  vs. MSSA 2–43 days,  $M_e=5.5$ ;  $p=0.038$ ).

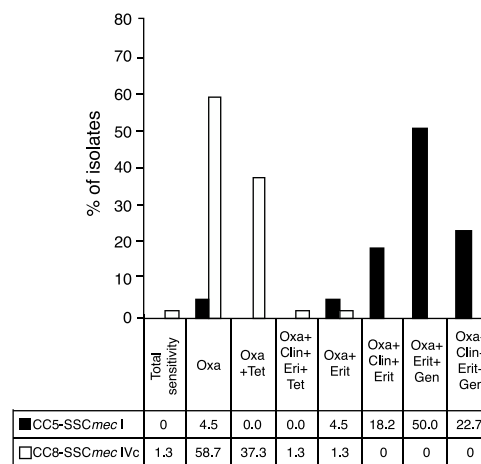
The most frequent sites of infection were skin and soft tissue, with 41.21% ( $n=75$ ), and the most common medical service was internal medicine, with 64.84% ( $n=110$ ).

The frequency of co-morbidities in the population was 74.1% ( $n=135$ ), more frequent in patients with POA infections caused by MSSA (MSSA 81.1%,  $n=30$  vs. MRSA 54.7%,  $n=35$ ;  $p=0.008$ ).

Of the population, 57.6% required surgical treatment and healing was the most frequent outcome; especially in POA infections caused by MSSA (MSSA 56.8%,  $n=21$  vs. MRSA 34.4%,  $n=22$ ;  $p=0.028$ ). Six of the patients included in the study died; however, this was the crude mortality and not attributed to *S. aureus* infection.

## Resistance profile

The MSSA isolates had six resistance profiles; 61.5% ( $n=40$ ) were susceptible to all antibiotics evaluated, 16.9% ( $n=11$ )



**Figure 2** Resistance profiles of the main MRSA clones. MRSA, methicillin-resistant *Staphylococcus aureus*. Figure shows the profile of antimicrobial resistance of the main MRSA clones found in the study. Antibiotics assessed: oxacillin (Oxa), tetracycline (Tet), clindamycin (Clin), erythromycin (Eri), gentamycin (Gen). Dark column: isolates belonging to CC5-SCCmec-I ( $n=22$ ); clear column: isolates belonging to CC8-SCCmec-IVc ( $n=75$ ).

were resistant to tetracycline, and 9.2% ( $n=6$ ) to erythromycin and clindamycin. The MRSA isolates had eight resistance profiles; 44.4% ( $n=52$ ) were resistant only to oxacillin, followed by 28.2% ( $n=33$ ) with resistance to oxacillin and tetracycline. All of the isolates were susceptible to vancomycin, linezolid, and tigecycline.

## Molecular typing

Among all the isolates, 16 clonal complexes (CC) were identified. MSSA strains were the more diverse, with a higher number of CCs; the most common were CC8 (29.2%,  $n=19$ ), CC45 (16.9%,  $n=11$ ), and CC1 (10.8%,  $n=7$ ), whereas in the MRSA strains the most common CCs were CC8 (70.9%,  $n=83$ ) and CC5 (22.2%,  $n=26$ ).

The presence of *mecA* gene was confirmed in the 117 selected MRSA strains. Of these, it was possible to identify the SCCmec type in 113 strains; four isolates were non-typeable. Of the MRSA isolates, 70.8% harbored SCCmec IVc ( $n=80$ ), followed by SCCmec I (20.3%,  $n=23$ ), SCCmec IVa (5.3%,  $n=6$ ), and SCCmec V (2.7%,  $n=3$ ). An isolate was identified with SCCmec IV, but its subtype was not identifiable (0.9%;  $n=1$ ).

Among patients with POA infections, 87.1% ( $n=54$ ) of MRSA isolates harbored SCCmec IVc, followed by SCCmec I (8.1%,  $n=5$ ) and SCCmec IVa (4.8%,  $n=3$ ). Meanwhile, MRSA isolates from HAI patients harbored SCCmec type IVc in 56.1% ( $n=23$ ), followed by SCCmec type I (26.8%;  $n=11$ ), IVa (7.3%;  $n=3$ ), V (7.3%,  $n=3$ ), and IV (2.43%,  $n=1$ ). SCCmec IVc was the most frequent type in both types of infection.

The resistance profiles of the most important MRSA clones are shown in Fig. 2.

Forty different *spa* types were identified in MSSA and MRSA strains. The most common types were t1610 (17.6%,  $n=32$ ), t008 (17%,  $n=31$ ), t149 (11%,  $n=20$ ), and t024 (9.9%;  $n=18$ ). In MSSA isolates belonging to CC8, types t1635 and

**Table 1** Clinical and epidemiological characteristics of patients with *Staphylococcus aureus* (MSSA–MRSA) infections.

	POA-I (n = 101) n (%)			HAI (n = 60) n (%)			Total (n = 182) n (%)		
	MSSA n = 37 (36.6)	MRSA n = 64 (63.4)	p-Value	MSSA n = 18 (30.0)	MRSA n = 42 (70.0)	p-Value	MSSA n = 65	MRSA n = 117	p-Value
<i>Gender</i>									
Male	25 (67.6)	41 (64.1)	0.721	11 (61.1)	32 (76.2)	0.235	43 (66.2)	78 (66.7)	0.944
Female	12 (32.4)	23 (35.9)		7 (38.9)	10 (23.8)		22 (33.8)	39 (33.3)	
<i>Age (years)</i>									
Median	4	4	0.534 <sup>b</sup>	3.5	1	0.141 <sup>b</sup>	3	2	0.330 <sup>b</sup>
Range	0–14	0–14		0–12	0–14		0–14	0–14	
≤ 1 years	10 (27.0)	21 (32.8)	0.601	5 (27.8)	22 (52.4)	0.130	23 (35.4)	52 (44.5)	0.826
2–4 years	12 (32.4)	16 (25.0)		5 (27.8)	10 (23.8)		18 (27.7)	27 (23.1)	
5–8 years	3 (8.1)	8 (12.5)		3 (16.7)	2 (4.8)		7 (10.8)	11 (9.5)	
9–12 years	7 (18.9)	15 (23.4)		5 (27.8)	5 (11.9)		12 (18.5)	20 (17.1)	
13–14 years	5 (13.5)	4 (6.3)		0 (0)	3 (7.1)		5 (7.7)	7 (6.0)	
<i>Previous history</i>									
Hospitalization in the past year	21 (56.8)	29 (45.3)	0.268	5 (27.8)	23 (54.8)	0.055	33 (50.8)	61 (52.1)	0.860
Stay in ICU in the past year	3 (8.1)	1 (1.6)	0.138 <sup>a</sup>	0 (0)	9 (21.4)	<b>0.047<sup>a</sup></b>	5 (7.7)	14 (12.0)	0.366
Surgery in the past year	11 (29.7)	11 (17.2)	0.141	4 (22.2)	14 (33.3)	0.389	25 (38.5)	36 (30.8)	0.292
Dialysis in the past year	4 (10.8)	1 (1.6)	0.059 <sup>a</sup>	1 (5.6)	4 (9.5)	1.000	5 (7.7)	5 (4.3)	0.333 <sup>a</sup>
MRSA isolation in the past year	1 (2.7)	2 (3.1)	1.000 <sup>a</sup>	0 (0)	4 (9.5)	0.306 <sup>a</sup>	1 (1.5)	7 (6.0)	0.262 <sup>a</sup>
Antimicrobials use in past six months	11 (31.4)	27 (45.8)	0.171	5 (29.4)	25 (64.1)	<b>0.017</b>	20 (32.8)	60 (55.0)	<b>0.005</b>
Trauma in past six months	9 (24.3)	10 (15.6)	0.281	7 (38.9)	8 (19.0)	0.118 <sup>a</sup>	16 (24.6)	19 (16.2)	0.169
Stay in a children's day care centers in the past year	1 (2.7)	6 (9.4)	0.418 <sup>a</sup>	0 (0)	5 (11.9)	0.309 <sup>a</sup>	1 (1.5)	13 (11.1)	<b>0.020</b>
Family infection in the past year	5 (21.7)	10 (21.7)	1.000	2 (15.4)	7 (19.4)	1.000 <sup>a</sup>	9 (21.4%)	17 (18.3)	0.668
Sports participation in the past year	8 (27.6)	16 (32.7)	0.639	2 (14.3)	3 (7.9)	0.602 <sup>a</sup>	10 (19.6)	20 (20.4)	0.908
<i>Hospital stay (days)</i>									
Median	0	0	0.098	5.50	14	<b>0.038</b>	0	0	0.309
Range	0–1	0–1	<sup>b</sup>	(2–43)	(0–434)	<sup>b</sup>	(0–50)	(0–434)	<sup>b</sup>

**Table 1** (Continued)

	POA-I (n = 101) n (%)			HAI (n = 60) n (%)			Total (n = 182) n (%)		
	MSSA n = 37 (36.6)	MRSA n = 64 (63.4)	p-Value	MSSA n = 18 (30.0)	MRSA n = 42 (70.0)	p-Value	MSSA n = 65	MRSA n = 117	p-Value
<i>Service</i>									
Internal medicine	28 (75.7)	42 (65.6)	0.264	10 (55.6)	27 (64.3)	0.369	43 (66.1)	75 (64.1)	0.388
Pediatric ICU	2 (5.4)	5 (7.8)		2 (11.1)	9 (21.4)		5 (7.7)	15 (12.8)	
Orthopedics	3 (8.1)	14 (21.9)		1 (5.6)	0 (0)		4 (6.2)	15 (12.8)	
Surgery	1 (2.7)	0 (0)		3 (16.7)	5 (11.9)		7 (10.8)	8 (6.8)	
Emergency	1 (2.7)	2 (3.1)		1 (5.6)	0 (0)		3 (4.6)	2 (1.7)	
Other	2 (5.4)	1 (1.6)		1 (5.6)	1 (2.4)		3 (4.6)	2 (1.7)	
<i>Infection type</i>									
Skin and soft tissue	18 (48.6)	32 (50.0)	0.011	9 (50)	16 (38.1)	0.329	27 (41.5)	48 (41.0)	0.155
Pneumonia	1 (2.7)	13 (20.3)		2 (11.1)	7 (16.7)		3 (4.6)	20 (17.1)	
Blood stream	4 (10.8)	5 (7.8)		4 (22.2)	3 (7.1)		8 (12.3)	8 (6.8)	
Catheter-related blood stream	4 (10.8)	0 (0)		3 (16.7)	11 (26.2)		7 (10.8)	11 (9.4)	
Osteomyelitis	1 (2.7)	6 (9.4)		0 (0)	0 (0)		1 (1.5)	6 (5.1)	
Arthritis	1 (2.7)	3 (4.7)		0 (0)	1 (2.4)		1 (1.5)	4 (3.4)	
Intra-abdominal	1 (2.7)	0 (0)		0 (0)	0 (0)		1 (1.5)	0 (0)	
Surgical site	-	-		-	-		10 (15.4)	11 (9.4)	
Other	7 (18.9)	5 (7.8)		0 (0)	4 (9.5)		7 (10.8)	9 (7.7)	
<i>Comorbidities</i>									
Co-morbidities	30 (81.1)	35 (54.7)	<b>0.008</b>	15 (83.3)	38 (90.5)	0.419	53 (81.5)	82 (70.1)	0.091
Atopy	7 (18.9)	8 (12.5)	0.382	1 (5.6)	8 (19.0)	0.255 <sup>a</sup>	9 (13.8)	16 (13.71)	0.974
Immunosuppression	6 (16.2)	5 (7.8)	0.204 <sup>a</sup>	3 (16.7)	3 (7.1)	0.352 <sup>a</sup>	9 (13.8)	9 (7.7)	0.183
Chronic renal disease	3 (8.1)	2 (3.1)	0.353 <sup>a</sup>	1 (5.6)	3 (7.1)	1.000 <sup>a</sup>	4 (6.2)	5 (4.3)	0.493 <sup>a</sup>
Neoplasia	3 (8.1)	1 (1.6)	0.138 <sup>a</sup>	0 (0)	2 (4.8)	1.000 <sup>a</sup>	3 (4.6)	4 (3.4)	0.702 <sup>a</sup>
Other co-morbidity <sup>c</sup>	12 (32.4)	13 (20.3)	0.174	6 (33.3)	24 (57.1)	0.091	26 (40.0)	45 (38.5)	0.838
Cardiovascular disease	3 (8.1)	1 (1.6)	0.138 <sup>a</sup>	1 (5.6)	5 (11.9)	0.658 <sup>a</sup>	11 (16.9)	11 (9.4)	0.136
Lung disease	1 (2.7)	1 (1.6)	1.000 <sup>a</sup>	0 (0)	8 (19.0)	0.091 <sup>a</sup>	1 (1.5)	9 (7.7)	0.099 <sup>a</sup>
<i>Treatment</i>									
Surgical treatment	22 (59.5)	43 (67.2)	0.435	8 (44.4)	20 (47.6)	0.821	34 (52.3)	71 (60.7)	0.273
<i>Outcome</i>									
Healing	21 (56.8)	22 (34.4)	<b>0.028</b>	14 (77.8)	35 (83.3)	0.719 <sup>a</sup>	41 (63.1)	62 (53.0)	0.188
Improvement	16 (43.2)	40 (62.5)	0.061	4 (22.2)	5 (11.9)	0.431 <sup>a</sup>	24 (36.9)	49 (41.9)	0.513
Death	0 (0)	2 (3.1) <sup>a</sup>	0.531 <sup>a</sup>	0 (0)	2 (4.8)	1.000 <sup>a</sup>	0 (0)	6 (5.1)	0.090 <sup>a</sup>

POA-I, present on admission infection; HAI, health care-associated infection; MSSA, methicillin-susceptible *Staphylococcus aureus*; MRSA, methicillin-resistant *Staphylococcus aureus*; ICU, intensive care unit.

Significant differences ( $p$ -values  $\leq 0.05$ ) are shown in bold.

<sup>a</sup> Fisher's exact test.

<sup>b</sup> Mann-Whitney  $U$ -test.

<sup>c</sup> Other co-morbidities: cardiovascular disease, chronic lung disease, nervous system diseases, skin diseases, malnutrition, cholestatic syndrome, subglottic stenosis, snake bite, Rh-ABO incompatibility, short bowel syndrome, nesidioblastosis, laryngeal and lung papillomatosis, hemophilia B, chronic osteomyelitis, congenital malformations, nephropathy, and others.

t008, each with 9.2% ( $n=6$ ) were the most frequent and t922, with 6.2% ( $n=4$ ), belonging to CC1, were highlighted. Most important strains of MRSA were those belonging to CC8, and the most frequent were CC8-SCCmec IVc-t1610 (27.43%,  $n=31$ ), CC8-SCCmec IVc-t008 (19.47%,  $n=22$ ), CC5-SCCmec I-t149 (16.81%,  $n=19$ ), and CC8-SCCmec IVc-t024 (15.04%,  $n=17$ ).

During the study period, a change was observed in the most important clonal complexes, both in MSSA and MRSA isolates. CC5 almost completely disappeared, while CC8 and CC45 remained constant. In addition, the increase of other CCs was evident, although none of them were specifically predominant. During the three years of the study, the clone belonging to CC8-SCCmec IVc of MRSA was increasing, representing the most prevalent, while the CC5-SCCmec I disappeared in the third year.

Sixteen isolates of MSSA were analyzed using PFGE, and a great diversity was found in the results (Fig. 3A). Analysis of MRSA by PFGE was performed on 26 isolates and four related groups were identified. The largest group had 15 isolates, which belonged to CC8, and harbored SCCmec IVc; 14 of them were positive for *pvl* and had different *spa* types (t008, 7/15; t1610, 4/15; t024, 3/15; t2031, 1/15; Fig. 3B).

### Virulence factors

Eight virulence factor genes were detected, both in MSSA and MRSA strains; however, higher gene prevalence was observed in MSSA compared to MRSA isolates. Statistically significant differences were observed between MSSA and MRSA strains regarding the *pvl* genes (MRSA 76.9%,  $n=90$  vs. MSSA 32.3%,  $n=21$ ;  $p=0.000$ ), *sed* (MSSA 33.8%,  $n=8$  vs. MRSA 10.3%,  $n=12$ ;  $p=0.000$ ), and *see* (MSSA 12.3%,  $n=8$  vs. MRSA 2.6%,  $n=3$ ;  $p=0.018$ ).

The presence of *pvl* showed statistically significant differences between isolates with SCCmec IVc and other types of SCCmec (SCCmec IVc 95%,  $n=76$  vs. other SCCmec 30.3%,  $n=10$ ;  $p=0.000$ ); It suggests that the *pvl* gene is more frequently associated with isolates with SCCmec IVc than SCCmec I. Genes such as *sed* and *tst* were associated with SCCmec IVa (*sed*,  $p=0.001$ ; *tst*,  $p=0.012$ ), while the *eta* gene was associated with SCCmec V ( $p=0.027$ ). No *etb* and *arcA* (ACME) genes were found in any of the isolates.

### Discussion

Infections caused by *S. aureus* (MSSA–MRSA) in child populations continue to be a major concern, both in the community and the hospital environment. In general, studies in the pediatric population are few, and some have limitations: many of them focus on certain infection types and most do not present information on infections caused by MSSA strains, which remain relevant and have not been displaced by MRSA strains.

In this study, a significant number of infections occurred on patients between birth and 1 year of age (41.2%), who mainly had skin and soft tissue infections caused by MRSA. Interestingly, few studies on *S. aureus* infections in children population agree with these findings, as reported in China by Wu et al. in 2010; they found that 37.6% of skin and soft

tissue infections occurred in children younger than 1 year.<sup>19</sup> Likewise, morbidity and mortality reports by the CDC in the United States have described an important prevalence of skin and soft tissue infections in neonatal patients.<sup>20</sup> However, other publications differ on infection types; authors such as Ilczyszyn et al. in Poland<sup>21</sup> and Qiao et al. in China<sup>22</sup> have shown that *S. aureus* infections in children between birth and 1 year are mainly of the invasive type (bacteremia and pneumonia).<sup>22</sup>

Likewise, results of other studies contrast with the present study regarding age; studies in Brazil<sup>23</sup> and Argentina,<sup>24</sup> for example, have shown a high prevalence of *S. aureus* infections in patients between 2 and 5 years of age, and other studies carried out in Colombia, in cities such as Bucaramanga<sup>25</sup> and Cartagena,<sup>5</sup> describe a high prevalence of *S. aureus* infections in children aged between 4 and 5 years and between 10 and 17 years, respectively.

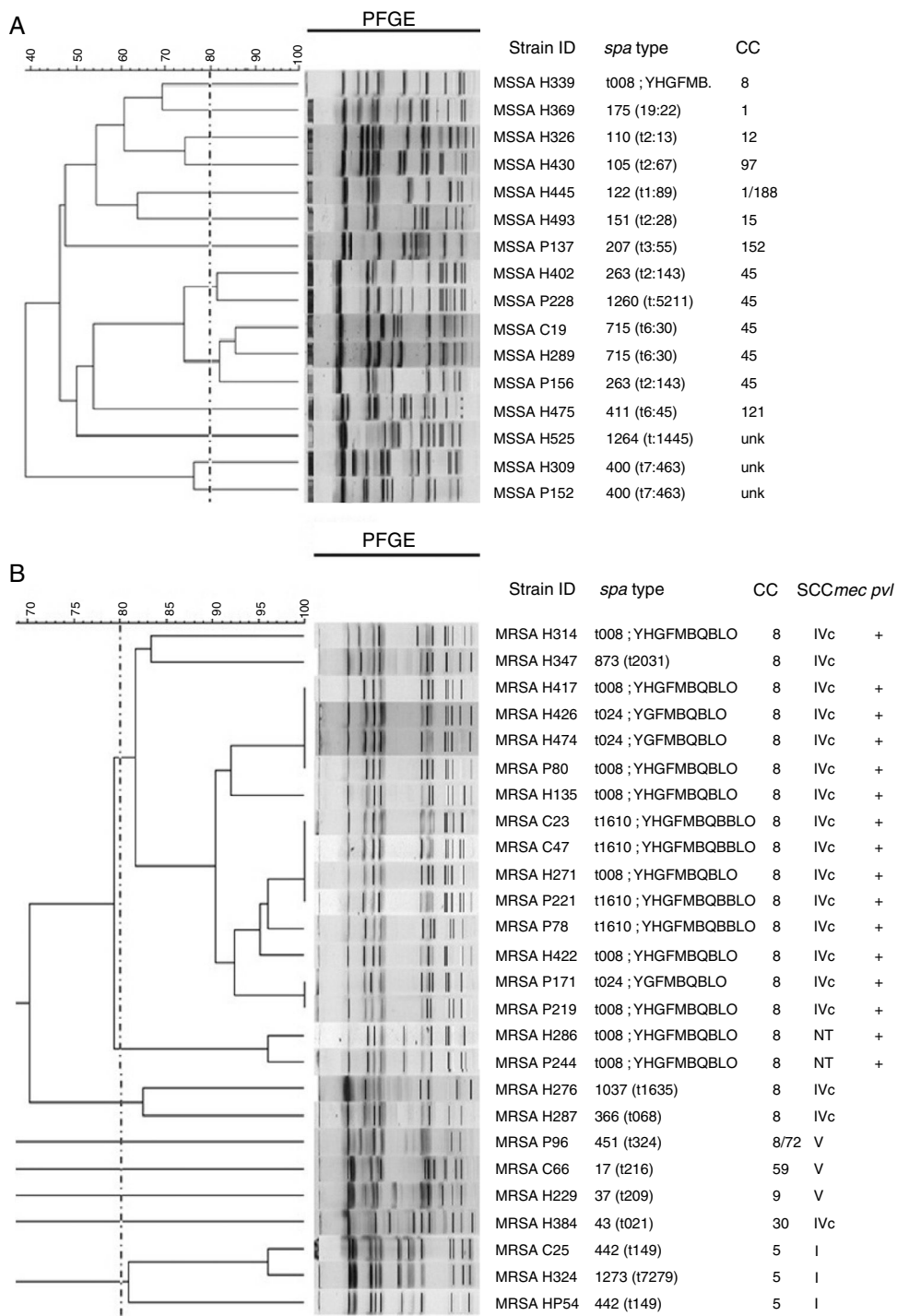
Infections in different age groups are determined by the particularities of each population and by the risk factors to which infants are exposed; furthermore, the infant population has a less adapted immune system, which allows greater colonization, more readily developed clinical presentations, and more complications.<sup>1,2</sup>

In this regard, a previous study, carried out in Medellín during 2011, revealed colonization frequencies by *S. aureus* (MSSA and MRSA) of 45.9% in children under 2 years.<sup>26</sup> This is an important aspect taking into account the relationship between colonization and the development of *S. aureus* infections, since it is possible that the frequencies of infection found in pediatric patients are related, among other aspects, to the frequencies of colonization reported in the city.

In particular, in this study a relationship between the antecedent of having stayed in day care centers and MRSA infections was found, which could be due to the frequency of colonization previously reported in children from Medellín day care centers. Rodríguez et al. were able to establish that the colonizing strains were closely related to the strains causing infection in the city.<sup>26</sup> Day care centers have been described as an important reservoir of the microorganism, becoming favorable places for its dissemination. These environments allow the swapping objects between children who have poor hygiene habits, which therefore makes dissemination more likely.<sup>2</sup>

According to a previous study published on *S. aureus* infections in the adult population from Medellín,<sup>27</sup> this study found that *S. aureus* strains harboring SCCmec IVc, usually associated with the community environment, predominated as etiological agents of HAIs, displacing the traditional strains with the SCCmec type I that dominated in hospitals. Further, frequencies of SCCmec IVc were higher in the infant population compared to those reported in the adult population (58.7% adult vs. 70.8% pediatric).<sup>27</sup>

This discovery shows the success of SCCmec IVc in the child population, which is a cause of great concern because it facilitates the maintaining of virulence factors like Pantone-Valentine leukocidin (PVL) and its dissemination can be greater in this population. As described, SCCmec IV is smaller; therefore, the biological cost of resistance decreases, favoring its propagation over other strains. Spread occurs mainly in strains belonging to specific clonal



**Figure 3** Genetic relatedness in MSSA and MRSA isolates. UPGMA dendrogram showing genetic relatedness in a sample of methicillin-susceptible *Staphylococcus aureus* (MSSA) (A) and methicillin-resistant *Staphylococcus aureus* (MRSA) (B) isolates. Dotted line indicates the cut-off point, or the Dice coefficient of 80%; this is used to define PFGE clones. Clusters above this percentage are considered to be genetically related. Isolates identified with the letter H belong to hospital A, isolates identified with the letter P belong to hospital B, and isolates identified with the letter C belong to hospital C.

complexes such as CC8, which could be evidenced in the PFGE results, as was reported previously.<sup>28</sup>

Nonetheless, the *spa* types predominant in this study; t1610, t008, t149, and t024 have been previously identified, mainly in MRSA strains infecting pediatric and adult

patients, not only in Colombia and Latin American but also in many countries around the world.<sup>29,30</sup> These findings show the wide circulation among the general populations, and suggest the pathogenic capacity and dissemination ability of these strains.



With regard to the resistance profiles, a difference was observed between this study and others, which may be due to differences in use of antimicrobials, either in the clinical setting or elsewhere. Heterogeneity confirms the importance of knowing the behavior of these infections in each region and institution. Further, the differences between the resistance profiles of the main MRSA clones reported in the study allow a clinical approach to the recognition of the predominant clones, without the need for molecular typing.

In the present study a predominance of POA infections were observed; however, health-care associated infections continue being very important in the pediatric population. At the same time, MSSA strains constitute an important source of infections; in this case, they harbored a variety of virulence factors genes in contrast with the MRSA strains, an aspect previously described.

The results of the present study represent valuable information for the knowledge of local epidemiology and the control of pediatric infections in the city. In addition, they demonstrate that the epidemiology of this microorganism is diverse and that, due to the particularities of each region, the data cannot be extrapolated. In general, the study demonstrated a higher prevalence of SCC*mec* IVc in children than adults, which could indicate a greater spread of the chromosomal cassette in this population group and may require additional studies.

Although the study group does not constitute a complete cohort, the patients and the evaluated isolates are the result of carefully designed sampling.

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## Conflicts of interest

The authors declare no conflicts of interest.

## References

- Rodríguez EA, Jiménez JN. Factores relacionados con la colonización por *Staphylococcus aureus*. IATREIA (Medellín). 2015;28:66–77.
- Lamaro-Cardoso J, De Lencastre H, Kipnis A, Pimenta FC, Oliveira LS, Oliveira RM, et al. Molecular epidemiology and risk factors for nasal carriage of *Staphylococcus aureus* and methicillin resistant *S. aureus* in infants attending day care center in Brazil. J Clin Microbiol. 2009;47:3991–7.
- Van der Mee Marquet N, Poisson DM, Lavigne JP, Francia T, Tristantan A, Vandenesch F, et al. The incidence of *Staphylococcus aureus* ST8-USA300 among French pediatric inpatients is rising. Eur J Clin Microbiol Infect Dis. 2015;34:935–42.
- Jimenez-Truque N, Saye EJ, Thomsen I, Herrera ML, Creech CB. Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in Costa Rican children. Pediatr Infect Dis J. 2014;33:e180–2.
- Correa-Jiménez O, Pinzón-Redondo H, Reyes N. High frequency of Pantón-Valentine leukocidin in *Staphylococcus aureus* causing pediatric infections in the city of Cartagena-Colombia. J Infect Public Health. 2016;9:415–20.
- Center for Disease Control and Prevention (CDC). Identifying healthcare-associated infections (HAI) for NHSN Surveillance; 2016. Available from: [https://www.cdc.gov/nhsn/pdfs/pscmanual/2psc\\_identifyinghais\\_nhsncurrent.pdf](https://www.cdc.gov/nhsn/pdfs/pscmanual/2psc_identifyinghais_nhsncurrent.pdf) [cited 05.04.16].
- Brakstad OG, Aasbakk K, Maeland JA. Detection of *Staphylococcus aureus* by polymerase chain reaction amplification of the *nuc* gene. J Clin Microbiol. 1992;30:1654–60.
- Mehrotra M, Wang G, Johnson WM. Multiplex PCR for detection of genes for *Staphylococcus aureus* enterotoxins, exfoliative toxins, toxic shock syndrome toxin 1, and methicillin resistance. J Clin Microbiol. 2000;38:1032–5.
- Shopsin B, Gomez M, Montgomery SO, Smith DH, Waddington M, Dodge DE, et al. Evaluation of protein A gene polymorphic region DNA sequencing for typing of *Staphylococcus aureus* strains. J Clin Microbiol. 1999;37:3556–63.
- Mathema B, Mediavilla J, Kreiswirth BN. Sequence analysis of the variable number tandem repeat in *Staphylococcus aureus* protein A gen: *spa* typing. Methods Mol Biol. 2008;431:285–305.
- Harmsen D, Claus H, Witte W, Rothgänger J, Claus H, Turnwald D, et al. Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for *spa* repeat determination and database management. J Clin Microbiol. 2003;41:5443–8.
- Enright MC, Day NP, Davies CE, Peacock SJ. Multilocus Sequence typing for characterization of methicillin-resistant methicillin-susceptible clones of *Staphylococcus aureus*. J Clin Microbiol. 2000;38:1008–15.
- Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. J Bacteriol. 2004;186:1518–30.
- Mulvey MR, Chui L, Ismail J, Louie L, Murphy C, Chang N, et al. Development of a Canadian standardized protocol for subtyping methicillin-resistant *Staphylococcus aureus* using pulsed-field gel electrophoresis. J Clin Microbiol. 2001;39:3481–5.
- Kondo Y, Ito T, Ma XX, Watanabe S, Kreiswirth BN, Etienne J, et al. Combination of multiplex PCRs for staphylococcal cassette chromosome *mec* type assignment: rapid identification system for *mec*, *ccr*, and major differences in junkyard regions. Antimicrob Agents Chemother. 2007;51:264–74.
- Milheirico C, Oliveira DC, de Lencastre H. Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome *mec* type IV in methicillin-resistant *Staphylococcus aureus*: “SCC*mec* IV multiplex”. J Antimicrob Chemother. 2007;60:42–8.
- Diep BA, Gill SR, Chang RF, Phan TH, Chen JH, Davidson MG, et al. Complete genome sequence of USA300, an epidemic clone of community-acquired methicillin-resistant *Staphylococcus aureus*. Lancet. 2006;367:731–9.
- McClure J, Conly JM, Lau V, Elsayed S, Louie T, Hutchins W, et al. Novel multiplex PCR assay for detection of the staphylococcal virulence marker Pantón-Valentine leukocidin genes and simultaneous discrimination of Methicillin-susceptible from resistant *Staphylococci*. J Clin Microbiol. 2006;44:1141–4.
- Wu D, Wang Q, Yang Y, Geng W, Wang Q, Yu S, et al. Epidemiology and molecular characteristics of community-associated methicillin-resistant and methicillin – susceptible *Staphylococcus aureus* from skin/soft tissue infections in a children’s hospital in Beijing, China. Diagn Microbiol Infect Dis. 2010;67:1–8.
- Center for Diseases Control and Prevention (CDC). Community-associated methicillin-resistant and *Staphylococcus aureus* infection among healthy newborns. Morb Mortal Wkly Rep. 2006;55:329–32.
- Ilczyszyn MW, Sabat AJ, Akkerboom V, Szkarlat A, Klepacka J, Sowa-Sierant I, et al. Clonal structure and characterization of

- Staphylococcus aureus* strains from invasive infections in pediatric patients from South Poland: association between age, *spa* types, clonal complexes, and genetic Markers. PLOS ONE. 2016;11:e0151937.
22. Qiao Y, Ning X, Chen Q, Zhao R, Song W, Zheng Y, et al. Clinical and molecular characteristics of invasive community-acquired *Staphylococcus aureus* infections in Chinese children. BMC Infect Dis. 2014;14:582.
  23. Gomes RT, Lyra TG, Alvez NN, Caldas RM, Barberino MG, Nascimento-Carvalho CM. Methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* infection among children. Braz J Infect Dis. 2013;17:573–8.
  24. Paganini H, Verdaguer V, Rodriguez AC, Della Latta P, Hernández C, Berberian G, et al. A clinical and risk factor analysis of methicillin-resistant community-acquired infections. Arch Argent Pediatr. 2006;104:295–300.
  25. Forero MJ. Infecciones por *Staphylococcus aureus* adquiridas en la comunidad. Diferenciación clínica según sensibilidad y resistencia a la meticilina en el servicio de infectología pediátrica del Hospital Universitario de Santander. 2006–2008. Santander: Universidad de Santander; 2009.
  26. Rodríguez EA, Correa MM, Ospina S, Atehortúa SL, Jiménez JN. Differences in epidemiological and molecular characteristics of nasal colonization with *Staphylococcus aureus* (MSSA–MRSA) in children from a university hospital and day care centers. PLoS ONE. 2014;9:e101417.
  27. Jiménez JN, Ocampo AM, Vanegas JM, Rodríguez EA, Mediavilla JR, Chen L, et al. CC8 MRSA strains harboring SCCmec type IVc are predominant in Colombian hospitals. PLoS ONE. 2012;7:e38576.
  28. Chambers HF, De Leo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. Nat Rev Microbiol. 2009;7:629–41.
  29. Machuca MA, Sosa LM, González CI. Molecular typing and virulence characteristic of methicillin-resistant *Staphylococcus aureus* isolates from pediatric patients in Bucaramanga, Colombia. PLoS ONE. 2013;8:e73434.
  30. Sola C, Paganini H, Egea AL, Moyano AJ, Garnerio A, Kevric I, et al. Spread of epidemic MRSA-ST5-IV clone encoding PVL as a major cause of community onset staphylococcal infections in Argentinean children. PLoS ONE. 2012;7:e30487.