



REVIEW ARTICLE

## What is new in genetics and osteogenesis imperfecta classification?☆



Eugênia R. Valadares<sup>a,\*</sup>, Túlio B. Carneiro<sup>a</sup>, Paula M. Santos<sup>b</sup>,  
Ana Cristina Oliveira<sup>b</sup>, Bernhard Zabel<sup>c</sup>

<sup>a</sup> Hospital das Clínicas, Faculdade de Medicina, Universidade Federal de Minas Gerais (UFMG), Belo Horizonte, MG, Brazil

<sup>b</sup> Faculdade de Odontologia, Universidade Federal de Minas Gerais (UFMG), Belo Horizonte, MG, Brazil

<sup>c</sup> Pediatric Clinic, Freiburg University, Freiburg, Germany

Received 17 March 2014; accepted 27 May 2014

Available online 18 July 2014

### KEYWORDS

Osteogenesis  
imperfecta;  
Osteochondrodysplasias;  
Collagen type 1

### Abstract

**Objective:** Literature review of new genes related to osteogenesis imperfecta (OI) and update of its classification.

**Sources:** Literature review in the PubMed and OMIM databases, followed by selection of relevant references.

**Summary of the findings:** In 1979, Sillence et al. developed a classification of OI subtypes based on clinical features and disease severity: OI type I, mild, common, with blue sclera; OI type II, perinatal lethal form; OI type III, severe and progressively deforming, with normal sclera; and OI type IV, moderate severity with normal sclera. Approximately 90% of individuals with OI are heterozygous for mutations in the *COL1A1* and *COL1A2* genes, with dominant pattern of inheritance or sporadic mutations. After 2006, mutations were identified in the *CRTAP*, *FKBP10*, *LEPRE1*, *PLOD2*, *PPIB*, *SERPINF1*, *SERPINH1*, *SP7*, *WNT1*, *BMP1*, and *TMEM38B* genes, associated with recessive OI and mutation in the *IFITM5* gene associated with dominant OI. Mutations in *PLS3* were recently identified in families with osteoporosis and fractures, with X-linked inheritance pattern. In addition to the genetic complexity of the molecular basis of OI, extensive phenotypic variability resulting from individual loci has also been documented.

**Conclusions:** Considering the discovery of new genes and limited genotype-phenotype correlation, the use of next-generation sequencing tools has become useful in molecular studies of OI cases. The recommendation of the Nosology Group of the International Society of Skeletal Dysplasias is to maintain the classification of Sillence as the prototypical form, universally accepted to classify the degree of severity in OI, while maintaining it free from direct molecular reference.

© 2014 Sociedade Brasileira de Pediatria. Published by Elsevier Editora Ltda.

Este é um artigo Open Access sob a licença de [CC BY-NC-ND](#)

☆ Please cite this article as: Valadares ER, Carneiro TB, Santos PM, Oliveira AC, Zabel B. What is new in genetics and osteogenesis imperfecta classification? J Pediatr (Rio J). 2014;90:536–41.

\* Corresponding author.

E-mail: [eugenia@medicina.ufmg.br](mailto:eugenia@medicina.ufmg.br), [eugeniavaladares@gmail.com](mailto:eugeniavaladares@gmail.com) (E.R. Valadares).

**PALAVRAS-CHAVE**

Osteogênese  
imperfeita;  
Osteocondrodisplasias;  
Colágeno tipo 1

**O que há de novo em genética e classificação de osteogênese imperfeita?****Resumo**

**Objetivo:** Revisão da literatura sobre novos genes relacionados à osteogênese imperfeita (OI) e atualização da sua classificação.

**Fonte dos dados:** Revisão nas bases de dados do PUBMED e OMIM com seleção de referências relevantes.

**Síntese dos dados:** Sillence *et al.*, em 1979, desenvolveram uma classificação dos subtipos de OI baseada em características clínicas e gravidade da doença: OI tipo I, forma leve, comum, com escleras azuladas; OI tipo II, forma perinatal letal; OI tipo III, forma grave e progressivamente deformante com esclera normal; e OI tipo IV, forma de gravidade moderada com esclera normal. Cerca de 90% dos indivíduos com OI são heterozigotos para mutações em *COL1A1* e *COL1A2*, com padrão de herança dominante ou esporádico. A partir de 2006 foram identificadas mutações nos genes *CRTAP*, *FKBP10*, *LEPRE1*, *PLOD2*, *PP1B*, *SERPINF1*, *SERPINH1*, *SP7*, *WNT1*, *BMP1* e *TMEM38B* associadas à OI recessiva e mutação em *IFITM5* associada à OI dominante. Mutações em *PLS3* foram identificadas recentemente em famílias com osteoporose e fraturas, com padrão de herança ligado ao X. Além da complexidade genética das bases moleculares das OI, extensa variabilidade fenotípica resultante de *loci* individuais também tem sido documentada.

**Conclusões:** Face à descoberta de novos genes e à correlação genótipo-fenótipo limitada, o uso de ferramentas de sequenciamento de nova geração torna-se útil no estudo molecular de casos de OI. A recomendação do Grupo de Nosologia da Sociedade Internacional de Displasias Esqueléticas é manter a classificação de Sillence como a forma prototípica e universalmente aceita para classificar o grau de gravidade na OI, e libertá-la de referência molecular direta.

© 2014 Sociedade Brasileira de Pediatria. Publicado por Elsevier Editora Ltda.

Este é um artigo Open Access sob a licença de [CC BY-NC-ND](#)

**Introduction**

Osteogenesis imperfecta (OI) is a group of clinically and genetically heterogeneous diseases characterized by susceptibility to bone fractures, with variable degree of severity and presumed or proven defects in collagen type I biosynthesis. Other manifestations include dentinogenesis imperfecta, blue sclerae, and short stature, as well as hearing loss in adulthood. Clinical manifestations range from severe cases with perinatal lethality to asymptomatic individuals with mild predisposition to fractures, normal stature, and normal life.<sup>1</sup>

Overall, the incidence of the different types of OI is approximately 1 in 15,000-20,000 births and most cases are due to autosomal dominant inheritance with mutations in *COL1A1* or *COL1A2* genes, which encode the  $\alpha 1$  (I) and  $\alpha 2$  (I) chains of type I collagen.<sup>1</sup>

Type I collagen, the main structural protein of the extracellular matrix of bone, skin, and tendons, consists of two pro- $\alpha$ -1 chains and one pro- $\alpha$ -2 chain that interweave, forming a rigid triple helix. Each  $\alpha$  chain contains N-(amino) and C-(carboxy) terminal propeptides and a central domain consisting of 338 repeats of Gly-XY, where X and Y exclude cysteine and tryptophan, and which often are, respectively, proline and hydroxyproline. Glycine, as the smallest amino acid, is the only residue that can occupy the axial position of the triple helix, so that any change in a glycine residue will result in the disruption of the helical structure.<sup>2,3</sup>

Mutations in *COL1A1* and *COL1A2* genes alter the structure or the amount of type I collagen, resulting in a skeletal phenotype that ranges from subclinical to lethal.<sup>1</sup>

These patients exhibit qualitative and quantitative abnormalities in type I collagen due to the dominant negative effect of the mutation, as the mutant pro- $\alpha$  chains are incorporated into the type I procollagen molecules that also contain normal pro- $\alpha$  chains. As a rule, when there is substitution of glycine in the  $\alpha 1$  chain, the phenotype will depend on the position of the substitution: C-terminal substitutions result in severe disease phenotype, and N-terminal substitutions yield milder phenotypes.<sup>4,5</sup> Residues with large lateral chains or charged residues are highly disruptive of the triple structure, regardless of where they are located. Different phenotypes have been found with the same mutation.<sup>6</sup>

In a consortium created in 2007 to study OI-causing mutations in type I collagen genes, 1,832 independent mutations were identified; 682 resulted in the substitution of glycine residues in the triple helix domain of the encoded protein, and 150 in splice sites.<sup>6</sup>

Based on clinical, radiographic, and skeletal findings, mode of inheritance, and molecular genetic analyses, new OI types have been identified since 2006 through exome sequencing. The present study aimed to review the classification of OI and to update new related genes. The PubMed and Online Mendelian Inheritance in Man (OMIM) databases were used.<sup>7</sup>

**Sillence Classification**

Due to considerable phenotypic variability, Sillence *et al.*<sup>8,9</sup> developed a classification of OI subtypes based on clinical features and disease severity (Table 1): OI type I, mild,

**Table 1** Classification of OI.<sup>a</sup>

Type	General manifestations	Specific manifestations
I- Autosomal dominant inheritance with blue sclera.	Variable bone fragility, blue sclera, early deafness, mild stunting.	I-A: normal teeth.  I-B and I-C: dentinogenesis imperfecta.
II- Perinatal lethal form, radiographically characterized by crumpled femora and beaded ribs.	Extreme bone fragility, perinatal death.	II-A: short and widened long bones with fractures, wide ribs with fractures.  II-B: short and widened long bones with fractures, ribs with sparse fractures. II-C: thin long bones with fractures, thin ribs.
III- Progressively deforming, with normal sclera.	Moderate to severe bone fragility, blue sclera in infancy.	Early-onset kyphoscoliosis.  Dentinogenesis imperfecta may be present.
IV- Autosomal dominant inheritance with normal sclera.	Bone fragility, moderate to severe deformity of the long bones and spinal column, white sclera, moderate to severe stunting.	IV-A: normal teeth.  IV-B: dentinogenesis imperfecta.

<sup>a</sup> Modified from Sillence et al.<sup>8</sup>

common, with blue sclera; OI type II, perinatal lethal form; OI type III, severe and progressively deforming, with normal sclera; and OI type IV, moderate severity with normal sclera. The classification of Sillence has been repeatedly revised when new causative genes for OI are identified.

### Expanded Classification

The molecular genetic classification of OI has shown to be very heterogeneous, with different patterns of inheritance and wide variability of clinical severity.<sup>10</sup>

Glorieux et al.<sup>11</sup> described an autosomal dominant form of OI, similar to OI Sillence type IV, but with distinct clinical, histological, and molecular characteristics. No mutations were found in *COL1A1* and *COL1A2* and, therefore, it was called OI type V (OMIM #610967) by the authors. Approximately 65% of affected individuals develop hyperplastic callus after fractures or surgical interventions, considered a pathognomonic characteristic.<sup>12</sup> Only in 2012 were *IFITM5* mutations identified in patients with type V OI, the gene encoding interferon-induced transmembrane protein 5, by sequencing of the entire exome.<sup>12-14</sup> The encoded protein has a role in early mineralization, but its mechanism remains unknown.<sup>10</sup>

In 2006, a *CRTAP* gene mutation was identified as the first genetic cause of lethal recessive OI.<sup>15</sup> Since then, new mutations in genes that cause recessive OI have been identified by exome sequencing, such as *FKBP10*, *LEPRE1*, *PLOD2*, *PP1B*, *SERPINF1*, *SERPINH1*, *SP*, *BMP1*, and *TMEM38B*. Each of these genes received an OI type number in the OMIM database, following the sequence numbers of the Sillence classification.

OI type VI (OMIM #613982) is an autosomal recessive form of the disease that can be caused by a homozygous mutation in the gene *SERPINF1* in chromosome 17p13.3, causing a mineralization defect.<sup>14</sup> According to the classification of Sillence et al., the phenotype is compatible with type IV or type III.<sup>16,17</sup>

OI type VII (MIM #610682) is a lethal autosomal recessive form of OI, caused by a mutation in *CRTAP* gene in homozygosity or compound heterozygosity in chromosome 3p22. It accounts for 2% to 3% of cases of lethal OI.<sup>15</sup>

Cabral et al.<sup>18</sup> described a form of autosomal recessive OI, called OI type VIII (OMIM #610915), which is characterized by white sclera, severe growth impairment, very poor skeletal mineralization, and bulbous metaphyses. This form is caused by mutations in the gene encoding leprecan (*LEPRE1*) in chromosome 1p34.2, associated with severe or lethal OI.

OI type IX (OMIM #259440) is an autosomal recessive form of OI corresponding to clinically severe types II/III of the Sillence classification.<sup>19</sup> There are no reports of dentinogenesis imperfecta. It can be caused by a homozygous mutation in the *PP1B* gene in chromosome 15q22.31.

OI type X (OMIM #613848) is an autosomal recessive form of the disease that can be caused by a homozygous mutation in the gene *SERPINH1* in chromosome 11q13.5. It is characterized by bone deformities and multiple fractures, generalized osteopenia, dentinogenesis imperfecta, and blue sclera. *SERPINH1* encodes a collagen-binding protein that acts as a chaperone in the endoplasmic reticulum and thus, individuals with mutations in this gene have cells that do not produce overmodified type I collagen.<sup>20</sup>

OI type XI (OMIM #610968) is an autosomal recessive form of the disease caused by a homozygous mutation in the *FKBP10* gene in chromosome 17q21, also related to a chaperone defect.<sup>1</sup> Patients with type OI type XI have severe progressive deformation and may have joint contractures. Patients do not have dentinogenesis imperfecta.<sup>21-23</sup>

OI type XII (OMIM #613849) is an autosomal recessive form, which can be caused by mutation in the *SP7* gene in chromosome 12q13.13. It is clinically characterized by recurrent fractures, mild bone deformities, generalized osteoporosis, delayed eruption of teeth, absence of dentinogenesis imperfecta, normal hearing, and white sclera.<sup>24</sup>

**Table 2** Expanded classification of OI.<sup>a</sup>

Type of OI	Inheritance	Phenotype	Genetic defect
<i>Classical Sillence Types</i>			
I	AD	Mild	<i>COL1A1</i>
	X-linked	Mild	<i>PLS3</i>
II	AD	Letal	<i>COL1A1</i> or <i>COL1A2</i>
III	AD	Progressive deformity	<i>COL1A1</i> or <i>COL1A2</i>
IV	AD	Moderate	<i>COL1A1</i> or <i>COL1A2</i>
V	AD	Moderate, hypertrophic callus and ossification of the interosseous membrane	<i>IFITM5</i>
VI	AR	Moderate to severe	<i>SERPINF1</i>
VII	AR	Severe to letal	<i>CRTAP</i>
VIII	AR	Severe to letal	<i>LEPRE1</i>
IX	AR	Severe to letal	<i>PPIB</i>
X	AR	Severe	<i>SERPINH1</i>
XI	AR	Progressive deformity, contractures	<i>FKBP10</i>
XII	AR	Moderate	<i>SP7</i>
XIII	AR	Severe	<i>BMP1</i>
XIV	AR	Variable severity	<i>TMEM38B</i>
XV	AR	Variable severity	<i>WNT1</i>
	AD	Early-onset osteoporosis	

AD, autosomal dominant; AR, autosomal recessive.

<sup>a</sup> Adapted from Forlino et al.<sup>1</sup>

OI type XIII (OMIM #614856) was described as caused by a homozygous mutation in the gene *BMP1* in chromosome 8p21.<sup>25,26</sup>

Shaheen et al.<sup>27</sup> described OI type XIV (OMIM # 615066), an autosomal recessive form characterized by varying degrees of severity with multiple fractures and osteopenia, with normal dentition, sclera, and hearing. Fractures occur prenatally or at approximately 6 years of age. It is caused by a homozygous mutation in the gene *TMEM38B* in chromosome 9q31.

OI type XV (OMIM #615220) has been designated based on the identification of mutations in *WNT1*.<sup>28–30</sup> Keupp et al.<sup>30</sup> reported that *WNT1* hypofunctional alleles result in phenotypes with low bone mass in humans. They verified that mutations in the recessive inherited gene lead to phenotypes of varying severity, ranging from mild to progressively deforming, which can occasionally lead to early infant death. They also detected families that had early osteoporosis with the autosomal dominant pattern of inheritance, with a heterozygous mutation in *WNT1*.

The recessive forms of OI with moderate to lethal phenotypes are caused by defects in genes whose products interact with collagen type I. Most recessive cases have null mutations in genes encoding proteins involved in prolyl 3-hydroxylation of collagen (*CRTAP*, *LEPRE1*, and *PPIB*), or in those responsible for the correct helical folding (*FKBP10* and *SERPINH1*). Types VII, VIII, and IX are caused by defects in 3-hydroxylation.<sup>1</sup> The genotype-phenotype correlation in recessive forms has been suggested.<sup>31</sup>

In 2013, *PLS3* mutations were identified in families with osteoporosis and fractures manifesting in childhood, with an X-linked pattern of inheritance.<sup>32</sup>

**Table 2** summarizes the classification based on the involved genes.

In 2010, van Dijk et al.<sup>33</sup> proposed a revised classification of OI, mentioning the causative gene and the corresponding clinical picture only for types I to VI. Types VII and VIII were excluded, as those types were added by genetic criteria, although their clinical and radiological findings were indistinguishable from those in types II to IV. The proposed classification leaves room for new genes discovered as the cause of OI until the full extent of heterogeneity is known.<sup>34</sup>

### OI Classification by the International Society of Skeletal Dysplasias

Due to the high genetic complexity of the molecular basis of OI and the extensive phenotypic variability resulting from individual loci described in recent years, it seemed impossible to maintain correlations between Sillence types and their molecular basis. But the proliferation of OI types to reflect each gene separately, supported by some, has become more confusing than useful in clinical practice. For these reasons, in 2009 the Nosology Group of the International Society of Skeletal Dysplasias recommended maintaining the classification of Sillence as the prototypical and universally accepted form to classify the degree of OI severity, and freeing it from direct molecular reference.<sup>35</sup> Thus, as shown in **Table 3**, OI was grouped into five clinical categories, and the several genes that can cause OI were listed separately. In the present study, the genes *IFITM5*, *SERPINF1*, *BMP1*, *WNT1*, *TMEM38B*, and *PLS3* were added to the original table, as they were discovered after its publication.

**Table 3** OI classification according to the International Society of Skeletal Dysplasias<sup>a</sup> with addition of newly discovered genes.

Osteogenesis Imperfecta	Inheritance	Genes
Nondeforming osteogenesis imperfecta (type I)	AD X-linked	<i>COL1A1</i> , <i>COL1A2</i> <i>PLS3</i>
Perinatal lethal (type II)	AD, AR	<i>COL1A1</i> , <i>COL1A2</i> , <i>CRTAP</i> , <i>LEPRE1</i> , <i>PPIB</i> , <i>BMP1</i>
Progressively deforming (type III)	AD, AR	<i>COL1A1</i> , <i>COL1A2</i> , <i>CRTAP</i> , <i>LEPRE1</i> , <i>PPIB</i> , <i>FKBP10</i> , <i>SERPINH1</i> , <i>SERPINF1</i> , <i>WNT1</i>
Moderate (type IV)	AD, AR	<i>COL1A1</i> , <i>COL1A2</i> , <i>CRTAP</i> , <i>FKBP10</i> , <i>SP7</i> , <i>SERPINF1</i> , <i>WNT1</i> , <i>TMEM38B</i>
With calcification of the interosseous membrane and/or hypertrophic callus (type V)	AD	<i>IFITM5</i>

AD, autosomal dominant; AR, autosomal recessive.

<sup>a</sup> Warman et al.<sup>35</sup>

## Conclusion

In practice, in spite of the complex genotypic variability of OI demonstrated in recent years, its phenotypes are still classified according to Sillence. Genotypic investigation should be indicated, especially in cases suggesting autosomal recessive inheritance, aimed at genetic counseling. The molecular study should be performed using Sanger sequencing of the several new genes, or by next-generation sequencing. Exome sequencing is useful when there is no panel of available genes, or when the involved genes are not known.

## Funding

CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico).

## Conflicts of interest

The authors declare no conflicts of interest.

## Acknowledgements

To the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) for the post-doctoral grant given to Eugênia Ribeiro Valadares in 2013 in the Genetics Sector of the Pediatric Clinic of Freiburg University, Germany, to develop the project "Investigation of osteogenesis imperfecta through the analysis of known genes and new candidate genes in Brazilian and German patients", under the supervision of Prof. Dr. Bernhard Zabel, Dr. Pablo Villavicencio Lorini, and Dr. Ekkehart Lausch, remarkable individuals.

## References

- Forlino A, Cabral WA, Barnes AM, Marini JC. New perspectives on osteogenesis imperfecta. *Nat Rev Endocrinol*. 2011;7:540–57.
- Cole WG. The molecular pathology of osteogenesis imperfecta. *Clin Orthop Relat Res*. 1997;343:235–48.
- Byers PH, Wallis GA, Willing MC. Osteogenesis imperfecta: translation of mutation to phenotype. *J Med Genet*. 1991;28:433–42.
- Stacey A, Bateman JF, Choi T. Perinatal lethal in transgenic mice bearing an engineered mutant pro-[alpha]1(I) collagen gene. *Nature*. 1988;332:131–6.
- Cole WG, Dagleis R. Perinatal lethal osteogenesis. *J Med Genet*. 1995;32:284–9.
- Marini JC, Forlino A, Cabral WA, Barnes AM, San Antonio JD, Milgrom S, et al. Consortium for osteogenesis imperfecta mutations in the helical domain of type I collagen: regions rich in lethal mutations align with collagen binding sites for integrins and proteoglycans. *Hum Mutat*. 2007;28:209–21.
- Online Mendelian Inheritance in Man, OMIM<sup>®</sup>. McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins University (Baltimore, MD), [cited 14 Mar 2014]. Available from: <http://omim.org/>
- Sillence DO, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. *J Med Genet*. 1979;16:101–16.
- Sillence DO. Osteogenesis imperfecta: an expanding panorama of variants. *Clin Orthop*. 1981;159:11–25.
- Marini JC, Blissett AR. New genes in bone development: what's new in osteogenesis imperfecta. *J Clin Endocrinol Metab*. 2013;98:3095–103.
- Glorieux FH, Rauch F, Plotkin H, Ward L, Travers R, Roughley P, et al. Type V osteogenesis imperfecta: a new form of brittle bone disease. *J Bone Miner Res*. 2000;15:1650–8.
- Semler O, Garbes L, Keupp K, Swan D, Zimmermann K, Becker J, et al. A mutation in the 5'-UTR of *IFITM5* creates an in-frame start codon and causes autosomal-dominant osteogenesis imperfecta type V with hyperplastic callus. *Am J Hum Genet*. 2012;91:349–57.
- Cho TJ, Lee KE, Lee SK, Song SJ, Kim KJ, Jeon D, et al. A single recurrent mutation in the 5'-UTR of *IFITM5* causes osteogenesis imperfecta type V. *Am J Hum Genet*. 2012;91:343–8.
- Balasubramanian M, Parker MJ, Dalton A, Giunta C, Lindert U, Peres LC, et al. Genotype-phenotype study in type V osteogenesis imperfecta. *Clin Dysmorphol*. 2013;22:93–101.
- Barnes AM, Chang W, Morello R, Cabral WA, Weis M, Eyre DR, et al. Deficiency of cartilage-associated protein in recessive lethal osteogenesis imperfecta. *N Engl J Med*. 2006;355:2757–64.
- Becker J, Semler O, Gilissen C, Li Y, Bolz HJ, Giunta C, et al. *Am J Hum Genet*. 2011;88:362–71.
- Glorieux FH, Ward LM, Rauch F, Lalic L, Roughley PJ, Travers R. Osteogenesis imperfecta type VI: a form of brittle bone disease with a mineralization defect. *J Bone Miner Res*. 2002;17:30–8.
- Cabral WA, Chang W, Barnes AM, Weis M, Scott MA, Leikin S, et al. Prolyl 3-hydroxylase 1 deficiency causes a recessive metabolic bone disorder resembling lethal/severe osteogenesis imperfecta. *Nature Genet*. 2007;39:359–65.
- van Dijk FS, Nesbitt IM, Zwikstra EH, Nikkels PG, Piersma SR, Fratantoni SA, et al. PPIB mutations cause severe osteogenesis imperfecta. *Am J Hum Genet*. 2009;85:521–7.
- Christiansen HE, Schwarze U, Pyott SM, AlSwaid A, Al Balwi M, Alrasheed S, et al. Homozygosity for a missense mutation in *SERPINH1*, which encodes the collagen chaperone protein HSP47,

- results in severe recessive osteogenesis imperfecta. *Am J Hum Genet.* 2010;86:389–98.
21. Alanay Y, Avaygan H, Camacho N, Utine GE, Boduroglu K, Aktas D, et al. Mutations in the gene encoding the RER protein FKBP65 cause autosomal-recessive osteogenesis imperfecta. *Am J Hum Genet.* 2010;86:551–9.
  22. Kelley BP, Malfait F, Bonafe L, Baldrige D, Homan E, Symoens S, et al. Mutations in FKBP10 cause recessive osteogenesis imperfecta and Bruck syndrome. *J Bone Miner Res.* 2011;26:666–72.
  23. Shaheen R, Al-Owain M, Faqeh E, Al-Hashmi N, Awaji A, Al-Zayed Z, et al. Mutations in FKBP10 cause both Bruck syndrome and isolated osteogenesis imperfecta in humans. *Am J Med Genet.* 2011;155A:1448–52.
  24. Lapunzina P, Aglan M, Temtamy S, Caparros-Martin JA, Valencia M, Leton R, et al. Identification of a frameshift mutation in Osterix in a patient with recessive osteogenesis imperfecta. *Am J Hum Genet.* 2010;87:110–4.
  25. Asharani PV, Keupp K, Semler O, Wang W, Li Y, Thiele H, et al. Attenuated BMP1 function compromises osteogenesis, leading to bone fragility in humans and zebrafish. *Am J Hum Genet.* 2012;90:661–74.
  26. Martinez-Glez V, Valencia M, Caparros-Martin JA, Aglan M, Temtamy S, Tenorio J, et al. Identification of a mutation causing deficient BMP1/mTLD proteolytic activity in autosomal recessive osteogenesis imperfecta. *Hum Mutat.* 2012;33:343–50.
  27. Shaheen R, Alazami AM, Alshammari MJ, Faqeh E, Alhashmi N, Mousa N, et al. Study of autosomal recessive osteogenesis imperfecta in Arabia reveals a novel locus defined by TMEM38B mutation. *J Med Genet.* 2012;49:630–5.
  28. Fahiminiya S, Majewski J, Mort J, Moffatt P, Glorieux FH, Rauch F. Mutations in WNT1 are a cause of osteogenesis imperfecta. *J Med Genet.* 2013;50:345–8.
  29. Pyott SM, Tran TT, Leistritz DF, Pepin MG, Mendelsohn NJ, Temme RT, et al. WNT1 Mutations in families affected by moderately severe and progressive recessive Osteogenesis Imperfecta. *Am J Hum Genet.* 2013;92:590–7.
  30. Keupp K, Beleggia F, Kayserili H, Barnes AM, Steiner M, Semler O, et al. Mutations in WNT1 cause different forms of bone fragility. *Am J Hum Genet.* 2013;92:565–74.
  31. Caparrós-Martin JA, Valencia M, Pulido V, Martínez-Glez V, Rueda-Arenas I, Amr K, et al. Clinical and molecular analysis in families with autosomal recessive osteogenesis imperfect identifies mutations in five genes and suggests genotype–phenotype correlations. *Am J Med Genet Part A.* 2013;161A:1354–69.
  32. van Dijk FS, Zillikens MC, Micha D, Riessland M, Marcelis CLM, Die-Smulders CE, et al. PLS3 mutations in X-linked osteoporosis with fractures. *N Engl J Med.* 2013;369:1529–36.
  33. van Dijk FS, Pals G, Van Rijn RR, Nikkels PG, Cobben JM. Classification of osteogenesis imperfecta revisited. *Eur J Med Genet.* 2010;53:1–5.
  34. van Dijk FS, Byers PH, Dagleish R, Malfait F, Maugeri A, Rohrbach M, et al. EMQN best practice guidelines for the laboratory diagnosis of osteogenesis imperfect. *Eur J Hum Genet.* 2012;20:11–9.
  35. Warman ML, Cormier-Daire V, Hall C, Krakow D, Lachman R, LeMerrer M, et al. Nosology and classification of genetic skeletal disorders: 2010 revision. *Am J Med Genet A.* 2011;155A:943–68.